

MOLECULAR



CURBING THE COVID-19 OUTBREAK IN REALTIME

# Abbott RealTime SARS-CoV-2

April 30<sup>th</sup>, 2020



**Abbott**

# Abbott's commitment to fight the COVID-19 pandemic

## Launched in the US and in EMEA:

- **RealTime SARS-CoV-2** assay for **m2000** received CE-IVD, US: FDA EUA. Scalable, automated process for flexible testing volumes (24-96 samples) and up to 470 patient samples in 24 hours.



## Launched in the US:

- **ID NOW SARS-CoV-2** assay EUA



## Launched in the US and in EMEA:

**SARS-CoV-2 IgG**



## In development:

- **Alinity m SARS-CoV-2**



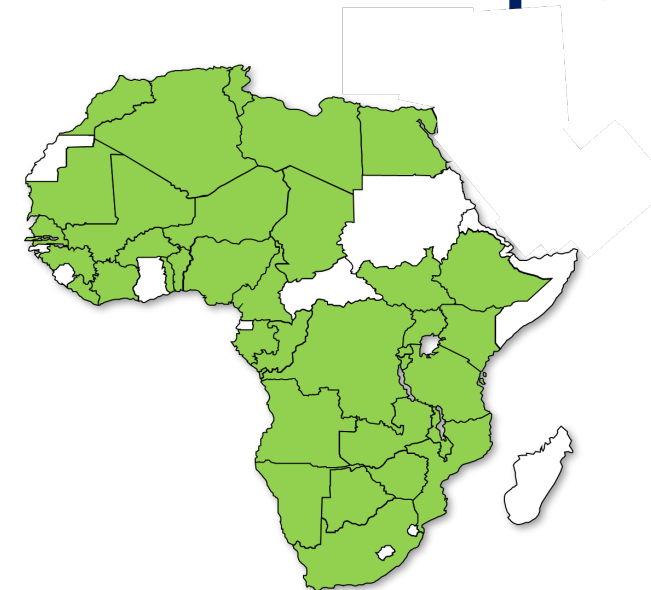
**Panbio COVID 19 IgG / IgM**



- **SARS-CoV-2 IgM**



## m2000 SYSTEM – Sub-Saharan Africa Placements & Capabilities



### Menu

- RealTime HIV-1 Viral Load
- RealTime HIV-1 Qualitative
- RealTime MTB
- RealTime MTB RIF/INH
- RealTime HCV Viral Load
- *maxCycle* HIV/HCV
- RealTime HCV Genotyping II
- RealTime HBV Viral Load
- RealTime CMV
- RealTime EBV

### Menu Cont

- RealTime High Risk HPV
- RealTime CT & CT/NG
- *m2000sp* open mode extraction capability
- *m2000rt* open mode capability
- **RealTime SARS-CoV-2**

Reliable (<2 calls/year)  
Efficient use of controls  
and floor space

# Abbott RealTime SARS-CoV-2 Specimen Types

## Nasal swab, Nasopharyngeal swab (NP) or Oropharyngeal swab (OP):

- Swab material: Sterile Dacron/nylon swab (Do not use calcium alginate swabs or swabs with wooden shafts, as they may contain substances that inactivate some viruses and inhibit PCR testing)
- Once sample is collected, the tip of the swab should be placed in a viral transport media tube (should contain 1-3mL of sterile viral transport medium)

### Common collection devices:

BD Universal Viral Transport Kits



COPAN UTM Viral Transport



Sources: <https://www.bd.com/en-us/offerings/capabilities/specimen-collection/swab-based-specimen-collection/bd-universal-viral-transport-system>;  
<https://www.copanusa.com/sample-collection-transport-processing/utm-viral-transport/>

# Abbott RealTime SARS-CoV-2 Sample Preparation

## Primary Tubes

- Collection devices may be loaded directly onto the *m2000sp*
- Swabs must be removed prior to loading
- Custom rack calibration available to minimize required dead volume

## Secondary Tubes

- Transfer 0.9-1.3mL of the viral/universal transport media from the collection device into either the *m2000sp* reaction vessel or transport tube
- Custom rack calibration available to minimize required dead volume

### Note:

- Laboratories should follow their own procedures for handling respiratory viruses before starting sample preparation on the *m2000sp*
- Please visit <https://www.cdc.gov/coronavirus/2019-nCoV/lab/lab-biosafety-guidelines.html> for biosafety guidance of SARS CoV-2 specimen handling

# Abbott RealTime SARS-CoV-2 Reagent Preparation

## Abbott *mSample Preparation System*<sub>DNA</sub> (LN 06K12-24)



- ***mLysis*<sub>DNA</sub>** - Add 35 mL ethanol to each bottle of *mLysis*<sub>DNA</sub>
- ***mMicroparticles*<sub>DNA</sub>**
- ***mWash1*<sub>DNA</sub>** - Add 23 mL ethanol to each bottle of *mWash1*<sub>DNA</sub>
- ***mWash2*<sub>DNA</sub>** - Add 70 mL ethanol to each bottle of *mWash2*<sub>DNA</sub>
- ***mElution*<sub>DNA</sub> Buffer**

# Abbott RealTime SARS-CoV-2 Dual Target Assay Design

- Dual Target, Single Stranded Linear Probes
- RdRp (RNA dependent RNA polymerase) and N-gene



CDC assay probes<sup>1</sup>

Charité Berlin / WHO<sup>2</sup>

Screening assay probes

Confirmation assay probes

Discriminatory assay probes

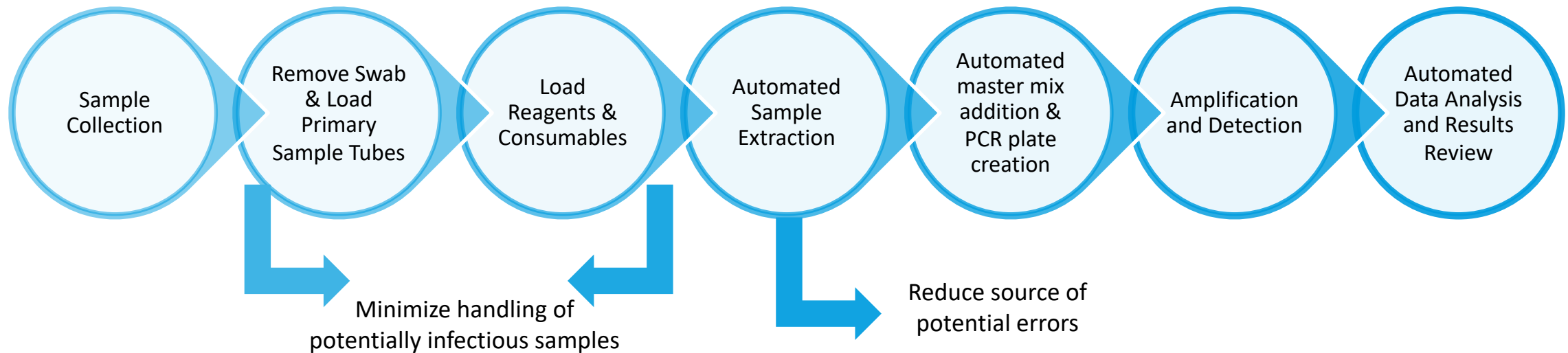
Abbott assay probes<sup>3</sup>



1. <https://www.cdc.gov/coronavirus/2019-ncov/downloads/rt-pcr-panel-for-detection-instructions.pdf> (accessed 7-Apr-2020)
2. <https://www.who.int/docs/default-source/coronaviruse/protocol-v2-1.pdf> (accessed 22-March-2020)
3. Abbott RealTime SARS-CoV-2 Assay PI: 51-608442/R1



# Automated Sample Handling Reduces Potential Sources of Error and Contamination



**Simple workflow with minimal sample handling  
Maintains 'Chain of Custody' and Provides confidence in results**



A clear interpretation of results enables laboratories to provide results to clinicians to quickly determine patient management and care

**Overview Orders Results System Help**

March 17, 2020 8:46 AM

**Abbott**

**Plate Results**

**Plates:**

Plate Name	Run Date and Time	Status	Archive Statu	Application Name
COVID PCs	3/16/2020 6:22 PM	Completed		0.6ml HIV-1 RNA
<b>TEST2</b>	<b>3/15/2020 7:20 PM</b>	<b>Completed</b>	<b>Archived</b>	<b>m2000_SARS-COV-2</b>
031220 AM1	3/12/2020 3:45 PM	Completed		COVID19_AM1 LDA
COVID31120	3/11/2020 4:19 PM	Completed		COVID19v1.2 LDA

**Results for Selected Plate:**

Location ▲	Sample Id	Sample Type	Assay	Result	Interpret	Flags	Error
A1	COV-2_NEG	Control	SARS-COV-2	Not Detected			
B1	COV-2_POS	Control	SARS-COV-2	24.18 CN			
C1	2		SARS-COV-2	17.48 CN	Positive		
D1	2		SARS-COV-2	24.19 CN	Positive		
<b>E1</b>	<b>2</b>		<b>SARS-COV-2</b>	<b>24.95 CN</b>	<b>Positive</b>		
F1	2		SARS-COV-2	24.85 CN	Positive		
G1	2		SARS-COV-2	24.99 CN	Positive		
H1	2		SARS-COV-2	24.89 CN	Positive		
A2	2		SARS-COV-2	25.55 CN	Positive		
B2	2		SARS-COV-2	25.21 CN	Positive		
C2	2		SARS-COV-2	25.73 CN	Positive		
D2	2		SARS-COV-2	25.06 CN	Positive		
E2	2		SARS-COV-2	25.17 CN	Positive		
F2	2		SARS-COV-2	25.25 CN	Positive		
G2	2		SARS-COV-2	25.88 CN	Positive		
H2	2		SARS-COV-2	24.68 CN	Positive		
A3	2		SARS-COV-2	Not Detected	Negative		
B3	2		SARS-COV-2	Not Detected	Negative		
C3	2		SARS-COV-2	Not Detected	Negative		
D3	2		SARS-COV-2	Not Detected	Negative		
E3	2		SARS-COV-2	Not Detected	Negative		
F3	2		SARS-COV-2	Not Detected	Negative		
G3	2		SARS-COV-2	Not Detected	Negative		
H3	2		SARS-COV-2	Not Detected	Negative		

**Plate Tasks**

- View Details
- View Result Details
- Release
- Archive
- Export
- Print Result List
- Print Result Details
- Print Errors
- Delete
- Analyze
- Export Results File

**Result Tasks**

- View Details
- Release
- Print List
- Print Details
- Delete

**m2000<sub>rt</sub>**

STOPPED Field Service Engineer: fse English

Overview Orders Results System Help

March 17, 2020 9:02 AM

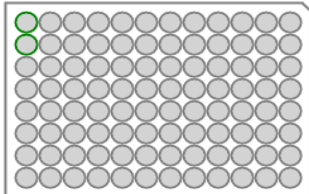
Abbott

## Plate Details

### Plate Information

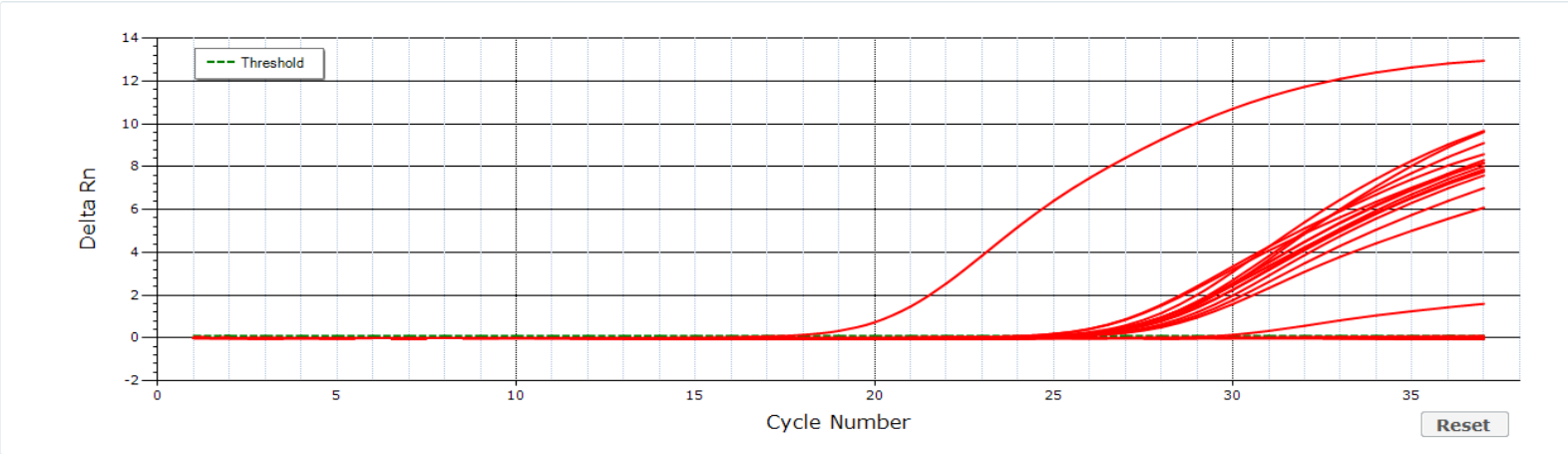
**Plate Name:** TEST2  
**Plate Status:** Completed  
**Run Completion Time:** 3/15/2020 7:20:34 PM  
**Amplification & Detection Application:** m2000\_SARS-COV-2 - 1.0  
**Operator:** fse  
**m2000sp Reagent Lot / Exp Date:** 10000658 / 5/31/2021  
**Plate Comment:**

### Graph Settings



- Calibrator
- Control
- Sample
- Empty Well

**Assay:** SARS-COV-2  
**Curve:** Target  
**Type:** Baselined  
**Y-Axis Scale:** Linear

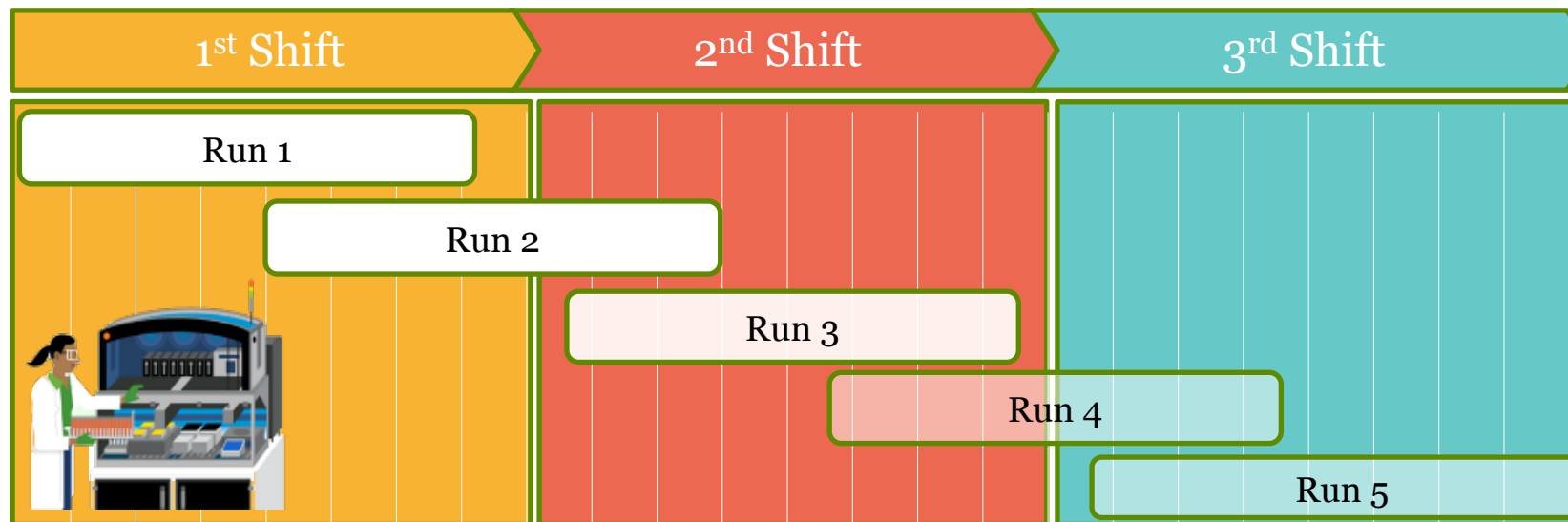


STOPPED Field Service Engineer: fse

Cancel Save

English

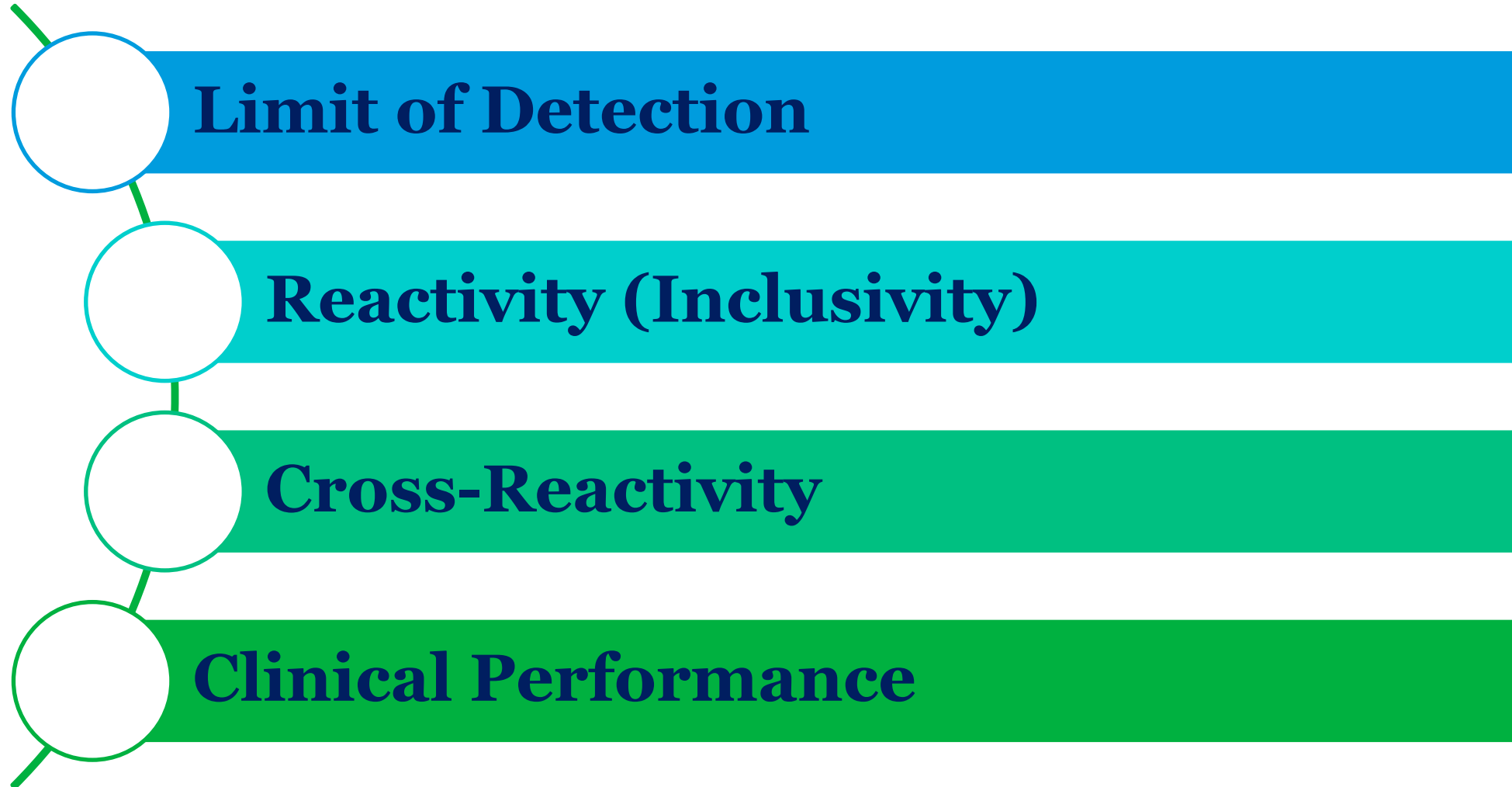
## Maximizing m2000sp/rt System Throughput



Abbott m2000 capacity of batches and tests per day			
With 1 m2000 system	up to 2 batches up to 188 tests	up to 3 batches up to 282 tests	up to 5 batches up to 470 tests
With 2 m2000 systems	up to 4 batches up to 376 tests	up to 6 batches up to 564 tests	up to 10 batches up to 940 tests
With 3 m2000 systems	up to 6 batches up to 564 tests	up to 9 batches up to 846 tests	up to 15 batches up to 1410 tests

Source: Abbott data on file

\*Run 1-5 = 94 tests + 2 Controls/Run\*



- Recombinant virus containing SARS-CoV-2 RNA was serially diluted in simulated nasal matrix (SNM).
- LOD was confirmed by testing 4 panel members with target concentrations at 400, 300, 200, and 100 copies/mL.
- LOD of the Abbott RealTime SARS-CoV-2 is 100 copies/mL with  $\geq 95\%$  detection

**Table 1.** LOD Determination Using Recombinant Virus Containing SARS-CoV-2

Virus Copies/mL	GE/Reaction <sup>1</sup>	Total Valid Replicates	Positive Replicates	Positive Rate (%)
400	12.5	21	21	100
300	9.4	21	21	100
200	6.2	21	21	100
100	3.1	21	20	95.2

<sup>1</sup>Genome equivalent per reaction (GE/reaction) was determined from calibration curve established using genomic RNA from SARS-Related Coronavirus 2, Isolate USAWA1/2020 (BEI Resources, Catalog No. NR-52285).

- Inclusivity was demonstrated by comparing the Abbott RealTime SARS-CoV-2 assay primers and probes to an alignment of all SARS-CoV-2 sequences available in Genbank as of March 5, 2020.
- The regions of the test's primers and probes were compared by in silico analysis to verify sequence homology with circulating SARS-CoV-2 strains.
- A total of 78 sequences from 10 countries (Australia, Belgium, Brazil, China, Finland, Nepal, South Korea, Sweden, Taiwan, and USA) had sequence coverage of at least one of the test's primers or probes for the comparison.
- Amongst the 78 sequences, there were also 6 strains without any country information listed in Genbank.

**All of the primers and probes in the test had 100% homology to all of the available circulating SARS-CoV-2 sequences.**

## In Silico Analysis

- Related pathogens, high prevalence disease agents, and normal or pathogenic flora that are reasonably likely to be encountered in the clinical specimen have been evaluated in silico to identify the % homology between the selected probe/primer sequences and the sequence present in the microorganism.
- *The conclusion of this analysis is that there is limited opportunity for cross-reactivity to allow for false-positive reporting or affect performance of SARS-CoV-2 virus detection.*



## Laboratory Testing

- Cross reactivity performance of Abbott RealTime SARS-CoV-2 assay was evaluated by testing 31 whole organisms or appropriate representative samples.
- *No cross-reactivity of the RealTime SARS-CoV-2 assay with the selected microorganisms was observed at the concentrations tested.*

Microorganism	Concentration	Result (No. Positive/No. Tested)	Final Result
Human coronavirus 229E	1.00 x 10 <sup>5</sup> Copies/mL	0/4	Negative
Human coronavirus OC43	1.00 x 10 <sup>5</sup> Copies/mL	0/4	Negative
Human coronavirus HKU1	Clinical Isolates	0/2	Negative
Human coronavirus NL63	1.00 x 10 <sup>5</sup> Copies/mL	0/4	Negative
SARS-coronavirus	25-28 (Ct range)	0/4	Negative
MERS-coronavirus	25-28 (Ct range)	0/4	Negative
Adenovirus (Ad. 71)	1.00 x 10 <sup>5</sup> TCID50/mL	0/4	Negative
Human Metapneumovirus (hMPV)	Clinical Isolates	0/3	Negative
Parainfluenza virus 1	1.00 x 10 <sup>5</sup> TCID50/mL	0/4	Negative
Parainfluenza virus 2	1.00 x 10 <sup>5</sup> Copies/mL	0/4	Negative
Parainfluenza virus 3	5.00 x 10 <sup>5</sup> TCID50/mL	0/4	Negative
Parainfluenza virus 4	Clinical Isolates	0/4	Negative
Influenza A (H1N1)	1.00 x 10 <sup>5</sup> Copies/mL	0/4	Negative
Influenza A / (H3N2)	1.00 x 10 <sup>5</sup> Copies/mL	0/4	Negative
Influenza B	1.00 x 10 <sup>5</sup> Copies/mL	0/4	Negative
Enterovirus Type 71	1.00 x 10 <sup>5</sup> TCID50/mL	0/4	Negative
Respiratory syncytial virus	1.00 x 10 <sup>5</sup> Copies/mL	0/4	Negative
Rhinovirus	1.00 x 10 <sup>5</sup> Copies/mL	0/4	Negative
<i>Chlamydia pneumoniae</i>	1.00 x 10 <sup>8</sup> IFU/mL	0/4	Negative
<i>Haemophilus influenzae</i>	1.00 x 10 <sup>8</sup> CFU/mL	0/4	Negative
<i>Legionella pneumophila</i>	1.00 x 10 <sup>8</sup> CFU/mL	0/3	Negative
<i>Mycobacterium tuberculosis</i>	1.00 x 10 <sup>8</sup> CFU/mL	0/4	Negative
<i>Streptococcus pneumoniae</i>	1.00 x 10 <sup>8</sup> CFU/mL	0/4	Negative
<i>Streptococcus pyogenes</i>	1.00 x 10 <sup>8</sup> CFU/mL	0/4	Negative
<i>Bordetella pertussis</i>	1.00 x 10 <sup>8</sup> CFU/mL	0/3	Negative
<i>Mycoplasma pneumoniae</i>	1.00 x 10 <sup>8</sup> CFU/mL	0/4	Negative
<i>Pneumocystis jirovecii (PJP)</i>	23-25 (Ct range)	0/4	Negative
<i>Candida albicans</i>	1.00 x 10 <sup>5</sup> CFU/mL	0/4	Negative
<i>Pseudomonas aeruginosa</i>	1.00 x 10 <sup>8</sup> CFU/mL	0/4	Negative
<i>Staphylococcus epidermis</i>	1.00 x 10 <sup>8</sup> CFU/mL	0/4	Negative
<i>S. salivarius</i>	1.00 x 10 <sup>8</sup> CFU/mL	0/4	Negative

- A clinical evaluation study was performed to evaluate the performance of the Abbott RealTime SARS-CoV-2 Assay using nasopharyngeal swab specimens.
- 61 contrived positive specimens at approximately 1X to 2X LOD and 20x LOD were tested. Samples were contrived by spiking known concentrations of recombinant virus containing SARS-CoV-2 RNA sequences into negative patient specimens.
- 34 negative specimens were tested.
- Positive and Negative Percent Agreement were 100%, respectively.

**Table 3. Clinical Evaluation of the Abbott RealTime SARS-CoV-2 Assay**

<b>SARS CoV-2 Concentration</b>	<b>Number Tested</b>	<b>Number Detected</b>	<b>% Detection</b>
1x to 2X LOD <sup>a</sup>	20	20	100
20X LOD	40	40	100
Negative <sup>b</sup>	31	0	0

<sup>a</sup> One replicate was invalid and excluded from the analysis; <sup>b</sup> Three replicates were invalid and excluded from the analysis.

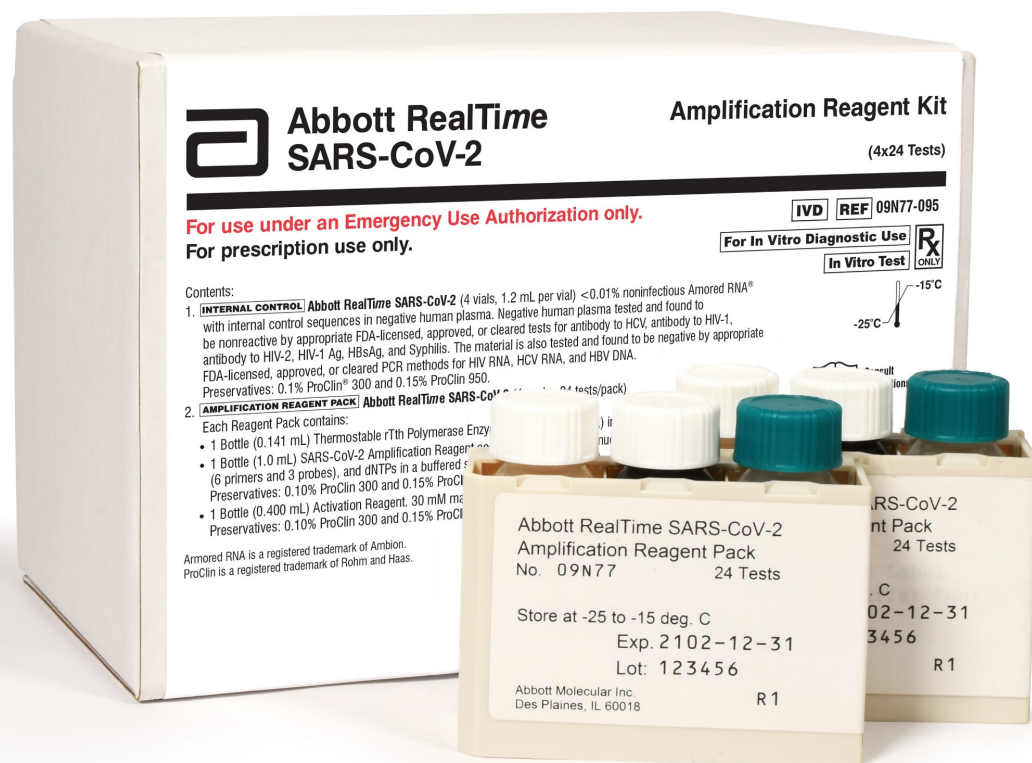
# Abbott RealTime SARS-CoV-2 Assay

Assay Specifications	
<b>Technology</b>	Qualitative Multiplex RT-PCR
<b>Probe Design</b>	Single Stranded Linear Probes
<b>Target Region</b>	Dual Target, RdRp and N-genes
<b>Assay Runtime*</b>	< 7 hours for 96 results
<b>Throughput*</b>	470 patient samples in 24 hours
<b>Specimen type</b>	Nasal, Nasopharyngeal and Oropharyngeal swabs
<b>Result Interpretation</b>	Positive / Negative
<b>Sample input volume</b>	0.5mL
<b>Internal Control (IC)</b>	Armored RNA (Pumpkin), Added to each specimen and control
<b>Controls</b>	One negative and One positive control per run



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# Abbott RealTime SARS-CoV-2 Assay



Dual Target

HIGHLY conserved target regions

Automated

MINIMIZE manual processes

Positive or Negative

SIMPLE & CLEAR result interpretation